

# Synthesis and Reactivity of Carbachlorins and Carbaporphyrins

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Supporting Information

ABSTRACT: Acid-catalyzed condensation of 3-ethoxymethylenecyclopentene-1-carbaldehyde with a tripyrrane, followed by oxidation with aqueous ferric chloride solutions, afforded moderate yields of a carbachlorin. This porphyrinoid exhibited a porphyrin-like UV-vis spectrum with a slightly intensified peak at 650 nm. The proton NMR spectrum showed that the carbachlorin is highly diatropic, and this has been confirmed by nucleus independent chemical shift (NICS) calculations. Oxidation of the carbachlorin with DDQ gave the first

example of a carbaporphyrin that is unsubstituted on the carbocyclic ring. Reaction of the carbachlorin with silver(I) acetate gave the corresponding silver(III) organometallic complex. When the carbachlorin was refluxed with methyl iodide and potassium carbonate in acetone, the 22-methyl derivative was formed. Treatment of the N-alkylation product with palladium(II) acetate afforded an unstable palladium(II) carbachlorin that was partially converted into a palladium(II) carbaporphyrin via an oxidation-methyl group migration process. Improved yields of the carbaporphyrin complex were obtained when the reaction mixture was stirred with aqueous ferric chloride solutions. These results open up the field of carbachlorin and carbaporphyrin chemistry for further study.

## INTRODUCTION

Carbaporphyrinoids are porphyrin analogues with at least one carbon atom within the macrocyclic cavity and include oxybenziporphyrins (1),<sup>2</sup> benzocarbaporphyrins (2),<sup>3</sup> azuliporphyrins (3),<sup>4</sup> and tropiporphyrins (4).<sup>5</sup> Oxybenziporphyrins and benzocarbaporphyrins retain porphyrin-like UV-vis spectra and are highly diatropic, as judged by proton NMR spectroscopy.<sup>6</sup> However, azuliporphyrins are cross-conjugated and have considerably reduced diatropic character, while tropiporphyrins have intermediary properties due to the nonplanar nature of the seven-membered ring. Chlorins (2,3dihydroporphyrins 5) are an important group of tetrapyrroles that provide the chromophore for many of the chlorophylls. Chlorins have modified UV-vis spectra and show relatively strong absorptions near 650 nm that lead to their green coloration in solution. There is a great deal of interest in porphyrinoid chromophores with strong absorptions at 650 nm or higher wavelengths, due in part to their potential application as photosensitizers in photodynamic therapy, and this has led to numerous investigations into the synthesis of chlorins. 9-13 However, there have been very few studies on the preparation of chlorin-like systems that may have equally useful properties. 14 Some time ago, we reported the synthesis of carbachlorins 6 using a "3 + 1" variant on the MacDonald condensation (Scheme 1). 15 Bicyclic dialdehydes 7 were found to condense with tripyrrane 8 in the presence of TFA to give, following oxidation with DDQ, propanocarbachlorin 6a and propenocarbachlorin 6b in moderate yields. These chlorin analogues retained aromatic properties, and the proton NMR spectra showed the inner CH protons upfield near -7 ppm. 15 The UV-vis spectra were also very porphyrin-like, showing a

# Scheme 1

Soret band at 404 nm and a series of small absorptions that culminated in a medium-sized Q band at 650 nm. 15 However, although true chlorins are easily oxidized to give porphyrins, attempts to further oxidize 6a,b to the corresponding carbaporphyrins 9 were unsuccessful.

In order to further explore the carbachlorin system, attempts were made to condense 1,3-cyclopentanedicarbaldehyde 10 with tripyrrane 8 to give porphyrinoid 11 (Scheme 2).<sup>15</sup>

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5 Chlorin

#### Scheme 2

However, no chlorinoid product could be isolated from these experiments. It was speculated that the presence of substituents, or the additional five-membered rings in structure 7, aids in the formation of porphyrinoid products by altering the conformation to the intermediates prior to cyclization. Indene dialdehyde 12 has been shown to be a particularly effective precursor to benzocarbaporphyrins 2,<sup>3</sup> and cyclopentadiene trialdehyde 13 has also been reacted with 8 to give carbaporphyrin aldehyde 14, albeit in low yields (Scheme 2).<sup>3,16</sup> The latter precursor is compromised by the presence of three aldehyde moieties, as only two of these are needed for

macrocycle formation and the third unit may react to give unwanted byproducts.

In relation to these earlier studies, it is notable that conjugated dialdehydes such as 12 are more effective in the "3 + 1" condensation reactions than aliphatic dialdehydes such as 7 and 10. Therefore, we speculated that the cyclopentene dialdehyde 15 might be a suitable precursor to chlorin 11 (Scheme 3). In this paper, the synthesis of carbachlorin 11 is

#### Scheme 3

$$\begin{array}{c}
Me \\
H \\
N \\
N \\
Et
\end{array}$$

$$\begin{array}{c}
\text{OHC} \\
\text{15}
\end{array}$$

described together with its conversion to a carbaporphyrin.<sup>17</sup> In addition, preliminary investigations into the metalation of these porphyrinoids are described.

## RESULTS AND DISCUSSION

Reports on the use of dialdehyde 15 in the synthesis of openchain conjugated systems have appeared in the patent literature, <sup>18</sup> but we could find no published methods for the synthesis of this compound. However, a convenient synthesis of the related acetal derivative 16 has been reported. <sup>19</sup> Reaction of cyclopentene-1-carbaldehyde with trimethylsilyl chloride and triethylamine gives the silyl dienol ether 17, and subsequent reaction with triethyl orthoformate and zinc chloride affords the related acetal 16 (Scheme 4). <sup>19</sup> Unfortunately, all attempts to

# Scheme 4

carry out an acid-catalyzed hydrolysis of 16 to give dialdehyde 15 were unsuccessful, either giving starting material or leading to decomposition. However, treatment of a chloroform solution of 16 with aqueous trifluoroacetic acid gave the related enol ether 18 instead. We reasoned that the reactivity of 18 should be similar to that of 15 and that this aldehyde could be used in the preparation of 11. As 18 proved to be rather unstable, a solution of the crude enol ether was generated and immediately used in reactions with tripyrrane 8 (Scheme 5). Tripyrrane 8 was treated with TFA and diluted with dichloromethane, and a freshly prepared solution of 18 in chloroform was added immediately. The mixture was allowed to react for 16 h and then oxidized by shaking the reaction mixture with 0.1% aqueous ferric chloride solution. Following purification by column chromatography and recrystallization, carbachlorin 11

## Scheme 5

was isolated in 11–16% yield. The proton and carbon-13 NMR spectra of 11 demonstrated that the porphyrinoid has a plane of symmetry and therefore must correspond to either tautomer 11 or a rapidly interconverting mixture of 11 and 11′ (Scheme 5). The proton NMR spectrum (Figure 1) also confirmed that this

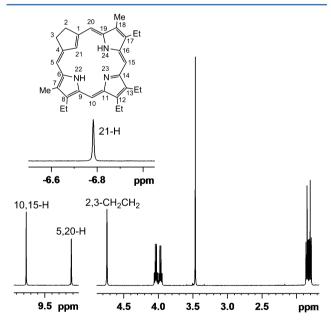
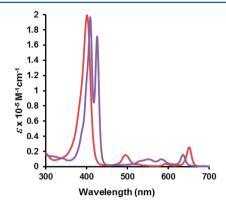


Figure 1. 500 MHz proton NMR spectrum of carbachlorin 11 in CDCl<sub>3</sub>.

porphyrinoid is highly diatropic, as the internal CH was observed at -6.8 ppm, while the external *meso* protons were shifted downfield to give two 2H singlets at 9.14 and 9.77 ppm. The CH<sub>2</sub>CH<sub>2</sub> unit of the carbachlorin ring gave a 4H singlet at 4.76 ppm due to its proximity to the macrocyclic ring current. The alkyl groups attached to the macrocycle are also strongly deshielded, and the methyl groups gave a 6H singlet at 3.47 ppm, while the CH<sub>2</sub> units of the four ethyl groups gave rise to two 4H quartets at 3.96 and 4.03 ppm. The carbon-13 NMR spectrum showed the internal CH at 122.1 ppm, while the *meso* 

carbons appeared at 96.2 (10,15-CH) and 97.4 ppm (5,20-CH). The UV-vis spectrum for 11 is very porphyrin-like, giving a Soret band at 401 nm and a series of Q bands at 495, 593, and 651 nm (Figure 2). As expected, the strongest of the

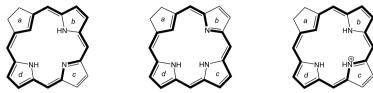


**Figure 2.** UV spectra of carbachlorin **11**: (red line) free base in 1% Et<sub>3</sub>N-99% CH<sub>2</sub>Cl<sub>2</sub>; (blue line) cation **11**H $^+$  in 1% TFA-99% CH<sub>2</sub>Cl<sub>2</sub>.

Q bands was the longest wavelength absorption. Addition of TFA afforded the cationic species 11H<sup>+</sup> (Scheme 5), and this gave a split Soret band at 409 and 426 nm (Figure 2). The UV–vis data closely resemble those previously reported for carbachlorins 6.<sup>15</sup> The proton NMR spectrum for cation 11H<sup>+</sup> in TFA–CDCl<sub>3</sub> demonstrated the presence of a slightly enhanced diatropic ring current. The inner CH gave a resonance at –6.95 ppm, while the NH protons afforded peaks at –5.52 (1H) and –4.19 ppm (2H). The external *meso* protons gave two 2H singlets at 9.53 and 10.02 ppm, while the carbachlorin methylene groups gave rise to a 4H singlet at 4.80 ppm. The proton and carbon-13 NMR spectra both confirmed that the cation retains a plane of symmetry.

In order to gain further insights into the carbachlorin system, the parent structure 19 was assessed using density functional theory calculations. 20 Chlorin tautomers 19 and 19' and the related cation 19H+ were minimized using B3LYP/6-311+ +G(d,p). Although the B3LYP functional has been shown to be reliable for calculating porphyrinoid geometries and frequencies, 21,22 the sensitivity of the relative energies to the choice of functionals needed to be assessed. 23,24 Single-point energy calculations were performed on the minimized structures using M06-2X/6-311++C(d,p) and B3LYP-D/6-311++G(d,p)(Table 1). 25,26 By these measures tautomer 19' is calculated to be between 4.79 and 5.00 kcal/mol less stable than tautomer 19. In 19, the NH units flank the imine nitrogen and are in a better position for hydrogen-bonding interactions. The carbachlorin structures are not completely planar, due to steric crowding within the macrocycle (Table 2). For 19, ring a is pivoted by nearly 10° relative to the mean macrocyclic plane, although the remaining rings are only slightly tilted (Table 2). Tautomer 19' is slightly more distorted, but this is magnified in cation 19H+, where four internal hydrogens have to be accommodated. The bond lengths in 19 are consistent with the presence of an  $18-\pi$ -electron delocalization pathway (Figure 3). The C6-C7, C8-C9, C16-C17, and C18-C19 bonds are relatively short in comparison to the C11-C12 and C13-C14 bonds, while the C7-C8 and C17-C18 bonds are longer than the C1--C13 bond, as would be predicted by the [18] annulene model for porphyrinoid aromaticity.<sup>27</sup> Nucleus independent

Table 1. Calculated Energies (hartrees), Relative Energies (kcal/mol), and NICS Values (ppm) for Tautomers of Carbachlorin 19 and the Related Cation 19H<sup>+</sup>.



	19	19'	<b>19</b> H <sup>+</sup>
B3LYP Minimum	-974.9472257	-974.9395878	-975.3458152
∆G (298 K)	0.00	4.59	***
<i>B3LYP/<b>B3LYP-D</b>/M06-2X</i>	0.00/ <b>0.00</b> /0.00	4.79/ <b>4.84</b> /5.00	***
NICS(0)	-14.03	-13.97	-13.95
NICS(a)	+5.81	+6.14	+6.07
NICS(b)	-14.02	-3.83	-14.61
NICS(c)	-5.11	-14.25	-15.84
NICS(d)	-14.02	-15.60	-14.61

Table 2. Angles (deg) between the Plane of Each of the Five-Membered Rings and the Overall Plane of the Macrocycle

molecule	ring a	ring $b$	ring c	ring d
19	11.10	1.97	1.26	1.99
19′	6.93	0.65	3.04	8.23
19H <sup>+</sup>	13.69	11.74	12.47	11.74

chemical shifts (NICS)<sup>28</sup> were also performed on 19, 19', and 19H<sup>+</sup>. The NICS(0) values for all three structures were close to -14 ppm, which is in accord with expectations for a highly diatropic porphyrinoid. The NICS values for the center of the individual subunits were also calculated. The pyrrole rings in 19 gave values of -14.02 ppm. The observed ring current is due to a combination of all of the possible delocalization pathways, but the  $18-\pi$ -electron pathway is likely to be dominant. As rings b and d lie within the aza[18]annulene pathway, large negative values would be expected. Nevertheless, the ring current due to the pyrrole ring itself may also be a significant factor. The pyrrolenine ring (c) gives a much smaller value of -5.11because the center of this ring lies outside of the  $18-\pi$ -electron delocalization pathway and this leads to deshielding. Ring a gives a value of +5.81, which is to be expected, as this ring also lies outside of the aza[18]annulene core and does not contribute a ring current of its own. Similar trends can be discerned for tautomer 19'. In cation 19H+, all three pyrrole rings give similar strongly negative values. This is due to

resonance contributors such as  $19'H^+$  and  $19''H^+$ , which facilitate three different versions of the aza[18]annulene pathway (Figure 4). However, ring *a* remains outside of these pathways, and this results in a deshielded NICS(a) value of +6.07 ppm.

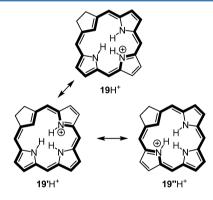


Figure 4. Alternative  $18-\pi$ -electron delocalization pathways in cation  $19\text{H}^+$ .

Attempts to oxidize carbachlorins **6** to the corresponding carbaporphyrins had been unsuccessful, but this may be due in part to the presence of a fused five-membered ring which would not favor the presence of sp<sup>2</sup>-hybridized carbons due to the associated angle strain.<sup>15</sup> When **11** was heated with DDQ in

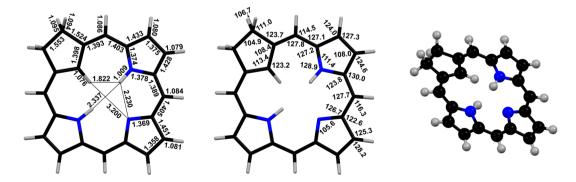


Figure 3. Bond lengths (Å) and angles (deg) and the 3D geometry for the minimized conformation of carbachlorin 19.

toluene, dehydrogenation readily occurred to give the corresponding carbaporphyrin 20 (Scheme 6). Most of the

#### Scheme 6

research on carbaporphyrins has made use of the readily available benzo-fused structures 2; therefore, access to 20 is an important advance in this area. Carbaporphyrin 11 is also highly diatropic and shows the internal CH as a singlet at -6.91 ppm, while the NHs produce a broad resonance at -3.92 ppm. The *meso* protons appear downfield as two 2H singlets at 9.77 and 9.83 ppm, and the external cyclopentadiene protons appear as a 2H doublet (J = 1.6 Hz) at 8.15 ppm (Figure 5). The

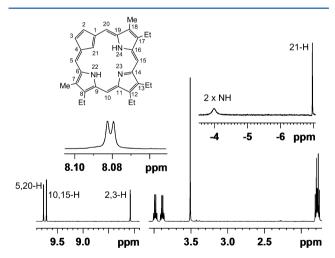
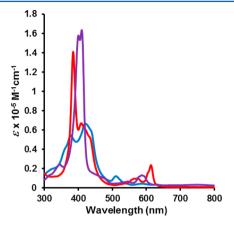


Figure 5. 500 MHz proton NMR spectrum of carbaporphyrin 20 in CDCl<sub>2</sub>.

doublet results from coupling to the inner CH (confirmed by  $^{1}\text{H}-^{1}\text{H}$  COSY), but this coupling is not resolved for the 21-H resonance. The proton and carbon-13 NMR spectra for **20** show that the macrocycle has a plane of symmetry, and this result is consistent with the depicted tautomer (the favorability of this tautomer is also supported by DFT calculations and is due in part to favorable hydrogen-bonding interactions within the macrocyclic cavity). The carbon-13 NMR spectrum in CDCl<sub>3</sub> showed the inner CH at 100.8 ppm, the *meso* carbons at 94.8 and 105.9 ppm, and the external cyclopentadiene carbons

at 128.7 ppm. The UV-vis spectrum of **20** was less porphyrinlike, showing two broad absorptions in the Soret region at 377 and 421 nm and broad Q bands at higher wavelengths (Figure 6). This is in contrast with the case for benzocarbaporphyrins **2** 



**Figure 6.** UV—vis spectra of carbaporphyrin **20**: (blue line) free base in 1% triethylamine—99% dichloromethane; (red line) monocation **20**H<sup>+</sup> with 10 equiv of TFA in CH<sub>2</sub>Cl<sub>2</sub>; (purple line) dication **20**H<sub>2</sub><sup>2+</sup> in 10% TFA—90% CH<sub>2</sub>Cl<sub>2</sub>.

and formylcarbaporphyrins 14, which show strong Soret bands and better defined Q bands in their UV-vis spectra. Addition of trace amounts of TFA produced a new species attributed to the monocation 20H<sup>+</sup>, which showed a strong Soret band at 385 nm, together with a broad absorption at 409 nm and Q bands at 563, 574, and 614 nm (Figure 6). Further addition of TFA gave rise to a third species corresponding to the Cprotonated dication 20H<sub>2</sub><sup>2+</sup>. In 10% TFA-90% CH<sub>2</sub>Cl<sub>2</sub>, a strong split Soret band appeared at 400 and 410 nm, together with weaker absorptions at higher wavelengths (Figure 6). Benzocarbaporphyrins 2 are also reported to form Cprotonated dications,<sup>3</sup> but diprotonation is only complete in approximately 50% TFA solutions, whereas diprotonation of 20 is essentially complete in 5% TFA solutions. The proton NMR spectrum of 20H<sub>2</sub><sup>2+</sup> shows the presence of a very strong diamagnetic ring current, and the internal CH2 gives a resonance at -8.27 ppm while the external cyclopentadiene protons appear at 11.11 ppm and the meso protons give rise to two 2H singlets at 11.00 and 11.45 ppm. These data show that the dication has enhanced diatropicity in comparison to the free base form. However, the downfield shift of the outer cyclopentadiene protons is due in part to the  $18-\pi$ -electron delocalization pathway being relocated through the fivemembered carbon ring (see bold pathway for 20H22+ in Scheme 6).

Carbaporphyrinoids such as 1, 2, and 4 react with silver(I) acetate to give silver(III) organometallic derivatives. <sup>32,33</sup> For instance, benzocarbaporphyrin 2 afforded good yields of the nonpolar silver(III) complexes 21 (Scheme 7). <sup>33</sup> This chemistry occurs rapidly in mixtures of dichloromethane and methanol at room temperature in the presence of 3 equiv of silver(I) acetate. However, carbachlorin 11 reacted rather slowly under these conditions. After 16 h, the silver(III) derivative 22 was isolated in 11% yield (Scheme 8). When the reaction was continued for several days, the yield was raised to 31%. However, when 7 equiv of silver(I) acetate was used, only low yields (6%) of impure silver(III) carbaporphyrin 23 could be isolated. Attempts to react carbaporphyrin 20 with silver(I)

#### Scheme 7

#### Scheme 8

acetate led to decomposition. These results indicate that the exposed carbocyclic ring is prone to oxidative degradation, and the presence of a fused benzene ring in 2 is clearly beneficial in stabilizing these structures. The UV—vis spectrum for 22 is porphyrin-like, showing a strong Soret band at 411 nm and a relatively strong Q band at 599 nm (Figure 7). The proton NMR spectrum for 22 indicates that the complex retains

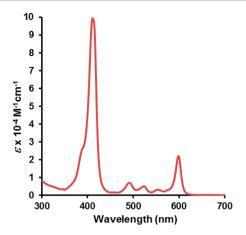


Figure 7. UV-vis spectrum of silver(III) carbachlorin 22 in dichloromethane.

strongly aromatic properties, as the *meso* protons show up downfield at 9.23 and 9.90 ppm and the carbachlorin  $\mathrm{CH_2CH_2}$  unit gives a 4H singlet at 5.02 ppm. The presence of a plane of symmetry was evident in the proton and carbon-13 NMR spectra, and the molecular formula was confirmed by high-resolution electron impact mass spectrometry.

Benzocarbaporphyrin does not give any isolatable products in reactions with palladium(II) salts. However, the N-alkyl derivatives 24 reacted with palladium(II) acetate in refluxing acetonitrile to give palladium(II) complexes 25 (Scheme 7). This chemistry is intriguing, because metalation is associated with an alkyl group migration from the nitrogen to the internal carbon atom.<sup>34</sup> A similar alkylation-metalation sequence was conducted on carbachlorin 11. Reaction of 11 with methyl iodide and potassium carbonate in refluxing acetone for 16 h gave the N-methyl derivative 26 in 34% yield. Prolonged reaction times did not improve the yield, as unidentified side products were formed. The site of alkylation can be deduced from the proton and carbon-13 NMR spectra, as the product shows a complete loss of symmetry (Figure 8). For instance, the meso protons now show up as four 1H singlets at 8.89, 9.04, 9.37, and 9.58 ppm. The alkylation introduces steric crowding within the macrocyclic cavity, but the NMR data show that the chlorin retains most of its diatropic character. The internal methyl group gave a 3H singlet at -4.27 ppm, while the inner CH gave a resonance at -6.31 ppm (Figure 8). As the methyl group is too large to pass through the cavity, carbachlorin 26 is chiral, and this leads to some complexity in the proton NMR spectrum. The 6 methylene units are all nonequivalent, and the individual protons are diastereotopic; thus, in principle this can lead to 12 different resonances. In reality there is a significant amount of overlap, but the carbachlorin CH<sub>2</sub>CH<sub>2</sub> unit gives rise to three multiplets at 4.40 (1H), 4.54 (1H), and 4.79 ppm (2H). The UV-vis spectrum for 26 in CH<sub>2</sub>Cl<sub>2</sub> is somewhat broadened in comparison to the spectrum for 11. The resulting weakened Soret band appeared at 407 nm, and less well-defined Q bands appeared between 500 and 664 nm (Figure 9). These differences presumably result from distortion of the carbachlorin chromophore due to the presence of the internal methyl group. Addition of TFA resulted in the formation of the cation 26H<sup>+</sup> with a split Soret band at 415 and 428 nm (Figure 9), and again the spectrum was somewhat broadened in comparison to 11H+. Methylated chlorin 26 was refluxed with palladium(II) acetate in acetonitrile for 30 min. Column chromatography on silica gel gave two fractions with similar

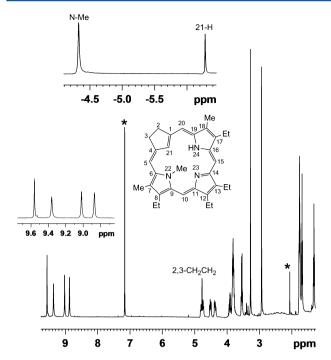
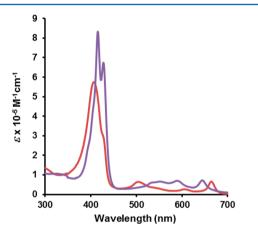


Figure 8. 500 MHz proton NMR spectrum of N-methylchlorin 26 in CDCl<sub>3</sub>. Asterisks denote solvent impurities.



**Figure 9.** UV—vis spectra of *N*-methylchlorin **26**: (red line) free base in 1% triethylamine—99% dichloromethane; (blue line) cation **26**H<sup>+</sup> in 1% TFA—99% CH<sub>2</sub>Cl<sub>2</sub>.

polarities corresponding to metalated products. It had been anticipated that the product of this chemistry would be the Nmethyl palladium(II) complex 27. Methyl group migration would not be expected in this case, as this would interrupt the macrocyclic conjugation pathway. Unexpectedly, the main product was identified as palladium(II) carbaporphyrin 28, where an oxidation had occurred in addition to the methyl group migration. Palladium complex 28 was isolated in 17% yield. The remaining fraction appeared to consist primarily of the expected palladium(II) chlorin 27, but this could not be isolated in pure form. The samples were always contaminated with 28, and it soon became apparent that 27 was slowly converting into 28. In order to speed up the oxidation, the crude reaction solution was diluted with dichloromethane and stirred vigorously with an aqueous ferric chloride solution for 24 h. Following workup, the yield of palladium(II) complex 28 was raised to 34%. It is likely that initial oxidation affords the N-

methyl palladium(II) carbaporphyrin 29 and this subsequently undergoes a rearrangement to give 28. The mechanism of the rearrangement is not known but could be a stepwise process involving the transient formation of a Pd-methyl bond. Alternatively, the conversion could be considered to be a [3,3]-sigmatropic shift.<sup>34</sup> Palladium complex 28 has regained a plane of symmetry and retains strongly diatropic characteristics. The proton NMR spectrum showed the internal methyl group at -4.46 ppm, while the meso protons appeared downfield as two 2H singlets at 10.00 and 10.42 ppm. The cyclopentadiene protons are directly connected to the  $18-\pi$ -electron delocalization pathway and as a result are deshelded to give a 2H singlet at 9.60 ppm. The UV-vis spectrum is less distinctive, giving broad bands in the Soret region at 394 and 440 nm. The molecular formula for 28 was confirmed by high-resolution electron impact mass spectrometry.

#### CONCLUSION

The synthesis of a carbachlorin using a "3 + 1" methodology has been accomplished, and this system retains highly aromatic characteristics. Oxidation with DDQ affords a related carbaporphyrin, and this also shows highly diatropic properties. The carbachlorin was metalated with silver(I) acetate to give a stable silver(III) organometallic derivative. In addition, the chlorin was reacted with methyl iodide to give a chiral N-methyl derivative. Reaction of the alkylated product with palladium(II) acetate afforded a palladium(II) complex via a metalation—oxidation—methyl group migration sequence. These results open this area up for the study of carbachlorins and carbaporphyrins without fused benzene units.

# **■ EXPERIMENTAL SECTION**

**General Considerations.** Melting points are uncorrected. NMR spectra were recorded using 400 and 500 MHz NMR spectrometers.  $^1\text{H}$  NMR values are reported as chemical shift δ, relative integral, multiplicity (s, singlet; d, doublet; t, triplet; q, quartet; m, multiplet; br, broad peak), and coupling constant (*J*). Chemical shifts are reported in parts per million (ppm) relative to CDCl<sub>3</sub> ( $^1\text{H}$  residual CHCl<sub>3</sub> δ 7.26,  $^{13}\text{C}$  CDCl<sub>3</sub> triplet δ 77.23 ppm), and coupling constants are taken directly from the spectra. NMR assignments were made with the aid of  $^1\text{H}-^1\text{H}$  COSY, HSQC, DEPT-135, and NOE difference proton NMR spectroscopy. 2D NMR experiments were performed using standard software. High-resolution mass spectra (HRMS) were carried out by using a double-focusing magnetic sector instrument.  $^1\text{H}$  and  $^{13}\text{C}$  NMR spectra for all new compounds are reported in the Supporting Information

**8,12,13,17-Tetraethyl-7,18-dimethylcarbachlorin** (11). Chloroform (20 mL) and a 50% aqueous trifluoroacetic acid solution (10 mL) were mixed, cooled to 0 °C, and then added to monoacetal **16** (200 mg, 1.31 mmol). The mixture was stirred at 0 °C for 3 h and then transferred to a separatory funnel. The organic layer was separated and the aqueous layer extracted with 30 mL of chloroform. The combined organic layers were washed with water, saturated sodium bicarbonate solution, and water. The organic layer was dried over sodium sulfate and filtered, and the resulting enol ether **18** solution was immediately used in the next step.

Tripyrranedicarboxylic acid 8<sup>35</sup> (200 mg, 0.47 mmol) was stirred with trifluoroacetic acid (4 mL) under nitrogen for several minutes. The solution was diluted with dichloromethane (200 mL), the aforementioned enol ether solution (approximately 100 mL) was added, and the resulting mixture was stirred under nitrogen at room temperature overnight. The solution was transferred to a separatory funnel and shaken with an aqueous ferric chloride solution (0.1%, 200 mL) for 5 min. The organic phase was separated and washed with water, saturated sodium bicarbonate solution, and water. The organic layer was dried over sodium sulfate and filtered, and the solvent was

removed under reduced pressure. The residue was chromatographed on a neutral grade 3 alumina column, with dichloromethane as eluent. The first fraction eluted as a purple solution which was evaporated under reduced pressure. The residue was further purified on a silica gel flash column, initially with dichloromethane and then with chloroform and 0.5% methanol-99.5% chloroform as eluents. A purple fraction was collected and recrystallized from chloroform-hexanes or chloroform-methanol to give the carbachlorin (21.2-31.9 mg, 0.047-0.070 mmol, 11-16%) as dark purple crystals: mp 246-247 °C; UV-vis (1% Et<sub>3</sub>N–99% CH<sub>2</sub>Cl<sub>2</sub>)  $\hat{\lambda}_{max}$  (log  $\varepsilon$ ) 401 (5.30), 495 (4.18), 526 (sh, 3.51), 593 (3.57), 651 nm (4.41); UV-vis (1% TFA-99% CH<sub>2</sub>Cl<sub>2</sub>)  $\lambda_{\text{max}}$  (log  $\varepsilon$ ) 409 (5.29), 426 (5.23), 530 (sh, 3.87), 552 (3.99), 582 (3.99), 636 nm (4.19); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  -6.77 (1H, s, 21-H), 1.80 (6H, t, J = 7.7 Hz, 12,13-CH<sub>2</sub>CH<sub>3</sub>), 1.84 (6H, t, J = 7.7Hz, 8,17-CH<sub>2</sub>CH<sub>3</sub>), 3.47 (6H, s, 7,18-CH<sub>3</sub>), 3.96 (4H, q, J = 7.7 Hz,  $12,13-CH_2$ ), 4.03 (4H, q, J = 7.7 Hz,  $12,13-CH_2$ ), 4.76 (4H, CH<sub>2</sub>CH<sub>2</sub>), 9.14 (2H, s, 5,20-H), 9.77 (2H, s, 10,15-H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  11.4 (7,18-CH<sub>3</sub>), 17.6 (8,17-CH<sub>2</sub>CH<sub>3</sub>), 18.9 (12,13-CH<sub>2</sub>CH<sub>3</sub>), 19.8 (8,17-CH<sub>2</sub>), 20.2 (12,13-CH<sub>2</sub>), 35.8 (2,3-CH<sub>2</sub>), 96.2 (10,15-CH), 97.4 (5,20-CH), 122.1 (21-CH), 128.7, 132.4, 136.1, 137.8, 142.7, 148.1, 149.5;  ${}^{1}$ H NMR (500 MHz, TFA-CDCl<sub>3</sub>)  $\delta$ -6.95 (1H, s, 21-H), -5.52 (1H, s, 24-NH), -4.19 2H, br s, 23,25-NH), 1.63 (6H, t, J = 7.7 Hz, 8,17-CH<sub>2</sub>CH<sub>3</sub>), 1.81 (6H, t, J = 7.7 Hz, 12,13-CH<sub>2</sub>CH<sub>3</sub>), 3.41 (6H, s, 7,18-CH<sub>3</sub>), 4.01 (4H, q, J = 7.7 Hz, 8,17- $CH_2$ ), 4.08 (4H, q, J = 7.7 Hz, 12,13- $CH_2$ ), 4.80 (4H, s,  $CH_2CH_2$ ), 9.53 (2H, s, 5,20-H), 10.02 (2H, s, 10,15-H); <sup>13</sup>C NMR (125 MHz, TFA-CDCl<sub>3</sub>)  $\delta$  11.6 (7,18-CH<sub>3</sub>), 16.8 (8,17-CH<sub>2</sub>CH<sub>3</sub>), 17.8 (12,13-CH<sub>2</sub>CH<sub>3</sub>), 19.9 (12,13-CH<sub>2</sub>), 20.1 (8,17-CH<sub>2</sub>), 36.3 (2,3-CH<sub>2</sub>), 93.6 (10,15-CH), 103.3 (5,20-CH), 131.0 (21-CH), 132.1, 133.6, 134.2, 139.5, 141.5, 142.6, 153.5; HR MS (EI) calcd for C<sub>31</sub>H<sub>37</sub>N<sub>3</sub> 451.2987, found 451.2992.

8,12,13,17-Tetraethyl-7,18-dimethylcarbaporphyrin (20). DDQ (10.0 mg, 0.044 mmol) was added to a solution of carbachlorin 11 (20.0 mg, 0.044 mmol) in toluene (10 mL). The mixture was stirred under reflux for 30 min and then cooled to room temperature. The solution was transferred to a separatory funnel, diluted with chloroform (50 mL), and washed with water. The aqueous layer was discarded, the organic layer was dried over sodium sulfate and filtered, and the solvent was removed under reduced pressure. The residue was chromatographed on grade 3 alumina, with 50% dichloromethane-50% hexanes as eluent. The first fraction (yellow) corresponded to carbaporphyrin 20. Recrystallization from chloroform-methanol gave **20** (14.2 mg, 0.0316 mmol, 71%) as dark green crystals: mp >300 °C; UV-vis (1% Et<sub>3</sub>N-99% CH<sub>2</sub>Cl<sub>2</sub>)  $\lambda_{\rm max}$  (log  $\varepsilon$ ) 377 (4.73), 421 (4.82), 510 (4.08), 585 nm (3.63); UV-vis (10 equiv TFA-CH<sub>2</sub>Cl<sub>2</sub>)  $\lambda_{\text{max}}$  $(\log \varepsilon)$  385 (5.15), 409 (4.83), 433 (sh, 4.72), 555 (sh, 3.91), 563 (3.98), 574 (4.00), 614 nm (4.37); UV-vis (10% TFA-90% CH<sub>2</sub>Cl<sub>2</sub>)  $\lambda_{\text{max}}$  (log  $\varepsilon$ ) 345 (4.39), 400 (5.20), 410 (5.21), 474 (sh, 4.01), 546 (3.83), 578 (sh, 4.07), 586 (4.12), 749 nm (3.51); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  -6.91 (1H, s, 21-H), -3.92 (2H, br s, 2 × NH), 1.84 (6H, t, J = 7.7 Hz, 8,17-CH<sub>2</sub>CH<sub>3</sub>), 1.87 (6H, t, J = 7.7 Hz, 12,13- $CH_2CH_3$ ), 3.58 (6H, s), 3.96, (4H, q, J = 7.7 Hz, 12,13- $CH_2$ ), 4.05  $(4H, q, I = 7.7 \text{ Hz}, 8.17 \text{-CH}_2), 8.15 (2H, d, I = 1.6 \text{ Hz}), 9.77 (2H, s, I)$ 10,15-H), 9.83 (2H, s, 5,20-H);  $^{13}$ C NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  11.5 (7,18-CH<sub>3</sub>), 17.6 (8,17-CH<sub>2</sub>CH<sub>3</sub>), 18.7 (12,13-CH<sub>2</sub>CH<sub>3</sub>), 19.7 (8,17-CH<sub>2</sub>), 20.1 (12,13-CH<sub>2</sub>), 94.8 (10,15-CH), 100.8 (21-CH), 105.9 (5,20-CH), 128.7 (2,3-CH), 133.4, 135.2, 136.6, 137.5, 138.2, 144.8, 153.8; <sup>1</sup>H NMR (500 MHz, TFA-CDCl<sub>3</sub>)  $\delta$  -8.27 (2H, s, 21-CH<sub>2</sub>), -4.42 (2H, br, 2 × NH), 1.78 (6H, t, I = 7.7 Hz, 12,13-CH<sub>2</sub>CH<sub>3</sub>), 1.82 (6H, t, J = 7.7 Hz, 8,17-CH<sub>2</sub>CH<sub>3</sub>), 3.76 (6H, s, 7,18-CH<sub>3</sub>), 4.21  $(4H, q, J = 7.7 \text{ Hz}, 8,17-\text{CH}_2), 4.26 (4H, q, J = 7.7 \text{ Hz}, 12,13-\text{CH}_2),$ 11.00 (2H, s, 10,15-H), 11.11 (2H, s, 2,3-H), 11.45 (2H, s, 5,20-H);  $^{13}$ C NMR (125 MHz, TFA-CDCl<sub>3</sub>)  $\delta$  11.9 (7,18-CH<sub>3</sub>), 16.6 (8,17-CH<sub>2</sub>CH<sub>3</sub>), 17.6 (12,13-CH<sub>2</sub>CH<sub>3</sub>), 20.36, 20.39 (8,12,13,17-CH<sub>2</sub>), 103.3 (10,15-CH), 113.6 (5,20-CH), 140.1, 141.4, 142.5, 146.3, 146.7, 147.6, 150.3 (2,3-CH), 151.9; HR MS (EI) calcd for C<sub>31</sub>H<sub>35</sub>N<sub>3</sub> 449.2831, found 449.2836.

[8,12,13,17-Tetraethyl-7,18-dimethylcarbachlorinato]silver-(III) (22). A solution of carbachlorin 11 (20.0 mg, 0.044 mmol) in dichloromethane (20 mL) was added to an suspension of 3.5 equiv of

silver(I) acetate (25.9 mg, 0.154 mol) in methanol (20 mL), and the mixture was stirred at room temperature for 4 days. The mixture was washed with water, and the solvent was removed under reduced pressure. The residue was purified by silica gel flash column, with dichloromethane as eluent. A green-pink fraction was collected and further recrystallized from chloroform—methanol to give the silver(III) carbachlorin (7.6 mg, 0.014 mmol, 31%) as a dark purple solid: mp >300 °C; UV-vis (CH<sub>2</sub>Cl<sub>2</sub>)  $\lambda_{\text{max}}$  (log  $\varepsilon$ ) 387 (sh, 4.40), 411 (5.00), 492 (3.85), 524 (3.71), 553 (3.49), 599 nm (4.34); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  1.82 (6H, t, J = 7.7 Hz, 8,17-CH<sub>2</sub>CH<sub>3</sub>), 1.88 (6H, t, J= 7.7 Hz, 12,13-CH<sub>2</sub>CH<sub>3</sub>), 3.46 (6H, s, 7,18-CH<sub>3</sub>), 4.00-4.05 (8H, m, 8,12,13,17-CH<sub>2</sub>), 5.02 (4H, s, CH<sub>2</sub>CH<sub>2</sub>), 9.23 (2H, s, 5,20-H), 9.90 (2H, s, 10,15-H);  $^{13}$ C NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  11.7 (7,18-CH<sub>3</sub>), 17.7 (8,17-CH<sub>2</sub>CH<sub>3</sub>), 18.5 (12,13-CH<sub>2</sub>CH<sub>3</sub>), 19.9, 20.5 (8,12,13,17-CH<sub>2</sub>), 36.3 (CH<sub>2</sub>CH<sub>2</sub>), 98.3 (10,15-CH), 100.5 (5,20-CH), 129.6, 133.7, 134.2, 135.5, 136.5, 138.1, 140.4; HR MS (EI) calcd for C<sub>31</sub>H<sub>34</sub>N<sub>3</sub>Ag 555.1804, found 555.1799.

8,12,13,17-Tetraethyl-7,18,22-trimethylcarbachlorin (26). Carbachlorin 11 (30.0 mg, 0.0665 mmol) was stirred with potassium carbonate (360 mg), 54 drops of iodomethane, and acetone (36 mL) under reflux for 16 h. The mixture was then cooled, diluted with dichloromethane (50 mL), and washed with water, and the solvent was removed under reduced pressure. The residue was run through a silica gel flash column with chloroform as eluent, and the starting material was initially collected as a dark green fraction. The major product fraction subsequently eluted, the solvent was removed under reduced pressure, and the residue was recrystallized from chloroformmethanol to yield the N-methylcarbachlorin (10.4 mg, 0.0224 mmol, 34%) as dark purple crystals: mp 173-174 °C; UV-vis (1% Et<sub>3</sub>N-99%  $CH_2Cl_2$ )  $\lambda_{max}$  (log  $\varepsilon$ ) 407 (4.76), 504 (3.81), 605 (3.42), 664 nm (3.82); UV–vis (1% TFA–99%  $CH_2Cl_2$ )  $\lambda_{max}$  (log  $\varepsilon$ ) 415 (4.92), 428 (4.83), 534 (sh, 3.79), 552 (3.82), 589 (3.85), 644 nm (3.86); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  -6.31 (1H, s, 21-H), -4.27 (3H, s, N- $CH_3$ ), -1.56 (1H, br s, NH), 1.38 (3H, t, J = 7.7 Hz), 1.77 (3H, t, J =7.7 Hz), 1.83–1.87 (6H, two overlapping triplets)  $(4 \times CH_2CH_3)$ , 2.94 (3H, s), 3.29 (3H, s) (7,18-CH<sub>3</sub>), 3.52-3.60 (2H, m), 3.78-3.86 (5H, m, 3.89-3.97 (1H, m) (4 × CH<sub>2</sub>CH<sub>3</sub>), 4.36-4.43 (1H, m), 4.51-4.56 (1H, m), 4.74-4.84 (2H, m) (CH<sub>2</sub>CH<sub>2</sub>), 8.89 (1H, s), 9.04 (1H, s), 9.37 (1H, s), 9.58 (1H, s)  $(4 \times meso-H)$ ; <sup>13</sup>C NMR (125) MHz, CDCl<sub>3</sub>)  $\delta$  11.55, 11.57, 16.3, 17.7, 18.55, 18.57, 19.82, 19.85, 20.0, 20.1, 30.9 (N-Me), 34.8, 36.4, 94.4, 98.2, 100.8, 101.7, 123.0, 128.3, 132.5, 132.7, 139.3, 140.1, 140.6, 141.2, 141.9, 143.5, 144.0, 145.7, 148.2, 148.5, 151.1;  $^{1}$ H NMR (500 MHz, TFA-CDCl<sub>3</sub>)  $\delta$  –7.09 (s, 1H), -6.18 (1H, s), -6.15 (1H, s) (2 × NH), -4.53 (3H, s, N- $CH_3$ ), 1.37 (3H, t, J = 7.7 Hz), 1.70 (3H, t, J = 7.7 Hz), 1.84–1.89 (6H, two overlapping triplets)  $(4 \times CH_2CH_3)$ , 3.13 (3H, s), 3.48 (3H, s) (7,18-CH<sub>3</sub>), 3.70-3.79 (2H, m), 4.00-4.09 (2H, m), 4.11-4.19 (4H, m)  $(4 \times CH_2CH_3)$ , 4.59-4.65 (1H, m), 4.67-4.72 (1H, m), 5.08-5.19 (2H, m) (CH<sub>2</sub>CH<sub>2</sub>), 9.50 (1H, s), 9.63 (1H, s), 10.05 (1H, s), 10.07 (1H, s) (4  $\times$  meso-H); HR MS (EI) calcd for  $C_{32}H_{39}N_3$ 465.3144, found 465.3136.

[8,12,13,17-Tetraethyl-7,18,21-trimethylcarbaporphyrinato]palladium(II) (28). Palladium(II) acetate (20 mg, 0.089 mmol) was added to a solution of N-methylcarbachlorin 26 (20.0 mg, 0.0430 mmol) in acetonitrile (20 mL), and the solution was stirred under reflux for 30 min. The mixture was cooled and diluted with chloroform (50 mL), and an aqueous solution of ferric chloride (0.1%, 60 mL) was added. The biphasic mixture was vigorously stirred at room temperature for 24 h. The two layers were separated, and the organic phase was washed with water and then evaporated to dryness. The residue was chromatographed with silica, with 30% dichloromethane-70% hexanes as eluent, and the product was collected as an emerald green fraction. Recrystallization from chloroform-methanol gave the palladium(II) complex (8.4 mg, 0.0148 mmol, 34%) as dark purple crystals: mp 229–230 °C; UV–vis (CH<sub>2</sub>Cl<sub>2</sub>)  $\lambda_{\rm max}$  (log  $\varepsilon$ ) 394 (4.48), 440 (4.33), 524 (3.74), 578 (sh, 3.76), 612 nm (3.86); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  -4.46 (3H, s, 21-CH<sub>3</sub>), 1.83-1.88 (12H, two overlapping triplets,  $4 \times CH_2CH_3$ ), 3.49 (6H, s, 7,18-CH<sub>3</sub>), 3.93-3.99 (8H, m, 4 × C $H_2$ C $H_3$ ), 9.60 (2H, s, 2,3-H), 10.00 (2H, s, 10,15-H), 10.42 (2H, s, 5,20-H);  $^{13}$ C NMR (125 MHz, CDC $l_3$ )  $\delta$  11.8 (7,18CH<sub>3</sub>), 17.8 (8,17-CH<sub>2</sub>CH<sub>3</sub>), 18.7 (12,13-CH<sub>2</sub>CH<sub>3</sub>), 19.8, 20.2 (4  $\times$  CH<sub>2</sub>CH<sub>3</sub>), 22.2 (21-CH<sub>3</sub>), 36.1, 102.3 (10,15-CH), 111.2 (5,20-CH), 133.9 (2,3-CH), 135.0, 141.6, 142.2, 142.9, 145.3, 148.0, 165.5; HR MS (EI) calcd for C<sub>32</sub>H<sub>35</sub>N<sub>3</sub>Pd 567.1866, found 567.1859.

**Computational Studies.** All calculations were performed using Gaussian  $09^{36}$  Rev D.01 running on a Linux-based PC. Energy minimization and frequency calculations of the porphyrinoid systems were performed at the density functional theory (DFT) level of theory with the B3LYP functional and a 6-311++G(d,p) basis set. Single-point energy calculations were performed on the minimized structures using both the B3LYP-D and M06-2X functionals with a 6-311++G(d,p) basis set. Mercury 3.1 running on an OS X platform, as provided by the CCDC (www.ccdc.cam.ac.uk/mercury/), was used to visualize the optimized structures. The resulting Cartesian coordinates of the molecules can be found in the Supporting Information.

NICS values were computed using the GIAO method,<sup>37</sup> at the DFT level of theory with the B3LYP functional and a 6-31+G(d,p) basis set, at several positions in each molecule. NICS(0) was calculated at the mean position of all the non-hydrogen atoms. NICS(a), NICS(b), NICS(c), and NICS(d) values were obtained by applying the same method to the mean positions of the non-hydrogen atoms that comprise the individual rings of each macrocycle.

## ASSOCIATED CONTENT

# **S** Supporting Information

Tables and figures giving Cartesian coordinates, HOMO and LUMO surfaces, and selected <sup>1</sup>H NMR, <sup>1</sup>H-<sup>1</sup>H COSY, HSQC, <sup>13</sup>C NMR, MS, and UV-vis spectra. This material is available free of charge via the Internet at http://pubs.acs.org.

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## Notes

The authors declare no competing financial interest.

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#### REFERENCES

- (1) (a) Lash, T. D. In Handbook of Porphyrin Science-With Applications to Chemistry, Physics, Material Science, Engineering, Biology and Medicine; Kadish, K. M., Smith, K. M., Guilard, R., Eds.; World Scientific Publishing: Singapore, 2012; Vol. 16, pp 1–329. (b) Pawlicki, M.; Latos-Grazynski, L. In Handbook of Porphyrin Science-With Applications to Chemistry, Physics, Material Science, Engineering, Biology and Medicine; Kadish, K. M., Smith, K. M., Guilard, R., Eds.; World Scientific: Singapore, 2010; Vol. 2, pp 104–192.
- (2) (a) Lash, T. D. Angew. Chem., Int. Ed. Engl. 1995, 34, 2533-2535.
  (b) Lash, T. D.; Chaney, S. T.; Richter, D. T. J. Org. Chem. 1998, 63, 9076-9088
- (3) (a) Lash, T. D.; Hayes, M. J. Angew. Chem., Int. Ed. Engl. 1997, 36, 840–842. (b) Lash, T. D.; Hayes, M. J.; Spence, J. D.; Muckey, M. A.; Ferrence, G. M.; Szczepura, L. F. J. Org. Chem. 2002, 67, 4860–4874
- (4) (a) Lash, T. D.; Chaney, S. T. Angew. Chem., Int. Ed. 1997, 36, 839–840. (b) Graham, S. R.; Colby, D. A.; Lash, T. D. Angew. Chem., Int. Ed. 2002, 41, 1371–1374. (c) Lash, T. D.; Colby, D. A.; Graham, S. R.; Chaney, S. T. J. Org. Chem. 2004, 69, 8851–8864.
- (5) (a) Lash, T. D.; Chaney, S. T. Tetrahedron Lett. 1996, 37, 8825–8828.
  (b) Bergman, K. M.; Ferrence, G. M.; Lash, T. D. J. Org. Chem. 2004, 69, 7888–7897.

- (6) Lash, T. D. Eur. J. Org. Chem. 2007, 5461-5481.
- (7) Scheer, H. In *Chlorophylls*; Scheer, H, Ed.; CRC Press: Boca Raton, FL, 1991; pp 3-30.
- (8) Pandey, R. K.; Zheng, G. In *Porphyrin Handbook*, Kadish, K. M., Smith, K. M., Guilard, R., Eds.; Academic Press: San Diego, CA, 2000; Vol. 6, pp 157–230.
- (9) Montforts, F. P.; Gerlach, B.; Höper, F. Chem. Rev. 1994, 94, 327-347.
- (10) Montforts, F.-P.; Glesenapp-Breiling, M. Prog. Heterocycl. Chem. 1998, 10, 1–24. For a recent review on the synthesis of bacteriochlorins and isobacteriochlorins, see: Brückner, C.; Samankumara, L.; Ogikubo, J. In Handbook of Porphyrin Science-With Applications to Chemistry, Physics, Material Science, Engineering, Biology and Medicine; Kadish, K. M., Smith, K. M., Guilard, R., Eds.; World Scientific: Singapore, 2012; Vol. 17, pp 1–112.
- (11) (a) Strachan, J.-P.; O'Shea, D. F.; Balasubramanian, T.; Lindsey, J. S. J. Org. Chem. 2000, 65, 3160–3172. (b) Taniguchi, M.; Ra, D.; Mo, G.; Balasubramaniam, T.; Lindsey, J. S. J. Org. Chem. 2001, 66, 7342–7354. (c) Taniguchi, M.; Kim, M. N.; Ra, D.; Lindsey, J. S. J. Org. Chem. 2005, 70, 275–285. (d) Muthiah, C.; Bhaumik, J.; Lindsey, J. S. J. Org. Chem. 2007, 72, 5839–5842. (e) Muthiah, C.; Ptaszek, M.; Nguyen, T. M.; Flack, K. M.; Lindsey, J. S. J. Org. Chem. 2007, 72, 7736–7749. (f) Mass, O.; Ptaszek, M.; Taniguchi, M.; Diers, J. R.; Kee, H. L.; Bocian, D. F.; Holten, D.; Lindsey, J. S. J. Org. Chem. 2009, 74, 5276–5289.
- (12) (a) Jacobi, P. A.; Lang, S.; Ghosh, I.; Leung, S. H.; Löwer, F.; Pippen, D. Org. Lett. **2001**, 3, 831–834. (b) O'Neal, W. G.; Roberts, W. P.; Ghosh, I.; Wang, H.; Jacobi, P. A. J. Org. Chem. **2006**, 71, 3472–3480. (c) O'Neal, W. G.; Jacobi, P. A. J. Am. Chem. Soc. **2008**, 130, 1102–1108.
- (13) (a) Burns, D. H.; Shi, D. C.; Lash, T. D. Chem. Commun. 2000, 299–300. (b) Burns, D. H.; Li, Y. H.; Shi, D. C.; Caldwell, T. M. J. Org. Chem. 2002, 67, 4536–4546. (c) Gryko, D. T.; Galezowski, M. Org. Lett. 2005, 7, 1749–1752. (d) Varamo, M.; Loock, B.; Maillard, P.; Grierson, D. S. Org. Lett. 2007, 9, 4689–4692. (e) Banerjee, S.; Zeller, M.; Brückner, C. J. Org. Chem. 2009, 74, 4283–4288.
- (14) (a) Callot, H. J.; Schaeffer, E. Tetrahedron 1978, 34, 2295–2300. (b) Vogel, E.; Kocher, M.; Balchi, M.; Teichler, I.; Lex, J.; Schmickler, H.; Ermer, O. Angew. Chem., Int. Ed. Engl. 1987, 36, 931–934. (c) McCarthy, J. R.; Jenkins, H. A.; Brückner, C. Org. Lett. 2003, 5, 19–22. (d) Lara, K. K.; Rinaldo, C. R.; Bruckner, C. Tetrahedron Lett. 2003, 44, 7793–7796. (e) Akhigbe, J.; Haskoor, J.; Krause, J. A.; Zeller, M.; Brückner, C. Org. Biomol. Chem. 2013, 11, 3616–3628. (f) Ogikubo, J.; Meehan, E.; Engle, J. T.; Ziegler, C. J.; Bruckner, C. J. Org. Chem. 2012, 77, 6199–6207.
- (15) Hayes, M. J.; Lash, T. D. Chem. Eur. J. 1998, 4, 508-511.
- (16) Berlin, K. Angew. Chem., Int. Ed. Engl. 1996, 35, 1820–1822.
- (17) These results were presented, in part, at the 246th ACS National Meeting, Indianapolis, IN, September 2013: Lash, T. D.; Li, D. Abstracts of Papers; American Chemical Society: Washington, DC, 2013; ORGN 322.
- (18) (a) Gross, W.; Oberkobusch, D.; Nemitz, R.; Hoeffkes, H. Ger. Offen. DE 102006042075 A1 20080306, 2008. (b) Maudauk, Y.; Payrastre, C.; Izquierdo, A. PCT Int. Appl. WO 2002046139 A2 20020613, 2002.
- (19) Makin, S. M.; Kruglikova, R. I.; Tagirov, T. K.; Kolobova, T. P. Zh. Org. Khim. 1983, 19, 1170–1174.
- (20) (a) Ghosh, A. Acc. Chem. Res. 1998, 31, 189–198. (b) Ghosh, A. In The Porphyrin Handbook; Kadish, K. M., Smith, K. M., Guilard, R., Eds.: Academic Press: San Diego, CA, 2000, Vol. 7, pp 1–38.
- (21) Alonso, M.; Geerlings, P.; De Proft, F. Phys. Chem. Chem. Phys. **2014**, *16*, 14396–14407.
- (22) Alonso, M.; Geerlings, P.; De Proft, F. Chem. Eur. J. 2013, 19, 1617–1628.
- (23) Schreiner, J. R. Angew. Chem., Int. Ed. 2007, 46, 4217-4219.
- (24) Wodrich, M. D.; Corminboeuf, C.; Schleyer, P. v. R. Org. Lett. **2006**, *8*, 3631–3634.
- (25) Zhao, Y.; Truhlar, D. G. Theor. Chem. Acc. 2008, 120, 215-241.
- (26) Grimme, S. J. Comput. Chem. 2004, 25, 1463-1473.

- (27) Lash, T. D. J. Porphyrins Phthalocyanines 2011, 15, 1093-1115.
- (28) Schleyer, P. v. R.; Maerker, C.; Dransfeld, A.; Jiao, H.; Hommes, N. J. R. v. E. *J. Am. Chem. Soc.* **1996**, *118*, 6317–6318.
- (29) The synthesis of a thiacarbachlorin and a related thiacarbaporphyrin has recently been reported: Berlicka, A.; Dutka, P.; Szterenberg, L.; Latos-Grazynski, L. *Angew. Chem., Int. Ed.* **2014**, 53, 4885–4889. However, it should be noted that benzothiacarbaporphyrins and related thiaoxybenziporphyrins were reported some time ago: Liu, D.; Ferrence, G. M.; Lash, T. D. *J. Org. Chem.* **2004**, 69, 6079–6093.
- (30) In our earlier studies on the metalation of benzocarbaporphyrins, <sup>32</sup> we referred to macrocycles such as **2** and **14** as "true carbaporphyrins" to differentiate them from other carbaporphyrinoid systems such as the N-confused porphyrins. Very recently, another group has used this term to refer to carbaporphyrins that do not have an additional fused ring. <sup>29</sup> In this publication, <sup>29</sup> it is not clear whether the authors even consider porphyrinoids **14a,b** to be "true" carbaporphyrins. However, by any measure **11** is a "true" carbachlorin and **20** is a "true" carbaporphyrin.
- (31) AbuSalim, D. A.; Lash, T. D. J. Org. Chem. 2013, 78, 11535–11548.
- (32) (a) Muckey, M. A.; Szczepura, L. F.; Ferrence, G. M.; Lash, T. D. *Inorg. Chem.* **2002**, *41*, 4840–4842. (b) Lash, T. D.; Colby, D. A.; Szczepura, L. F. *Inorg. Chem.* **2004**, *43*, 5258–5267. (c) Lash, T. D.; Rasmussen, J. M.; Bergman, K. M.; Colby, D. A. *Org. Lett.* **2004**, *6*, 549–552.
- (33) Furuta, H.; Ogawa, T.; Uwatoko, Y.; Araki, K. Inorg. Chem. 1999, 38, 2676-2682.
- (34) Lash, T. D. Org. Lett. 2011, 13, 4632-4635.
- (35) (a) Sessler, J. L.; Johnson, M. R.; Lynch, V. J. Org. Chem. 1987, 52, 4394–4397. (b) Lash, T. D. J. Porphyrins Phthalocyanines 1997, 1, 29–44.
- (36) Frisch, M. J.; Trucks, G. W.; Schlegel, H. B.; Scuseria, G. E.; Robb, M. A.; Cheeseman, J. R.; Scalmani, G.; Barone, V.; Mennucci, B.; Petersson, G. A.; Nakatsuji, H.; Caricato, M.; Li, X.; Hratchian, H. P.; Izmaylov, A. F.; Bloino, J.; Zheng, G.; Sonnenberg, J. L.; Hada, M.; Ehara, M.; Toyota, K.; Fukuda, R.; Hasegawa, J.; Ishida, M.; Nakajima, T.; Honda, Y.; Kitao, O.; Nakai, H.; Vreven, T.; Montgomery, J. A., Jr.; Peralta, J. E.; Ogliaro, F.; Bearpark, M.; Heyd, J. J.; Brothers, E.; Kudin, K. N.; Staroverov, V. N.; Kobayashi, R.; Normand, J.; Raghavachari, K.; Rendell, A.; Burant, J. C.; Iyengar, S. S.; Tomasi, J.; Cossi, M.; Rega, N.; Millam, N. J.; Klene, M.; Knox, J. E.; Cross, J. B.; Bakken, V.; Adamo, C.; Jaramillo, J.; Gomperts, R.; Stratmann, R. E.; Yazyev, O.; Austin, A. J.; Cammi, R.; Pomelli, C.; Ochterski, J. W.; Martin, R. L.; Morokuma, K.; Zakrzewski, V. G.; Voth, G. A.; Salvador, P.; Dannenberg, J. J.; Dapprich, S.; Daniels, A. D.; Farkas, Ö.; Foresman, J. B.; Ortiz, J. V.; Cioslowski, J.; Fox, D. J. Gaussian 09, Revision D.01; Gaussian, Inc., Wallingford, CT, 2009.
- (37) Wolinski, K.; Hinton, J. F.; Pulay, P. J. Am. Chem. Soc. 1990, 112, 8251–8260.